

The Effects of Different Levels of Rosehip Fruit Added in the Rations of Laying Hens Raised Under High Altitude and Cold Stress on Some Blood Parameters, Rectal Temperature, Fertility Rate and Chick Quality

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Abstract

This study was carried out to determine the effects of different doses (5g/kg, 10g/kg, 15g/kg) of rosehip fruit, which is a source of ascorbic acid (vitamine C), on fertility rate, chick quality and some blood parameters in the feed of breeder hens and roosters raised under high altitude and cold stress. It was applied during 12-week trial. While the fertility rate was not significantly affected by the treatments, the plasma vitamine C content of the laying hens was significantly affected by the treatments ($P < 0.0001$). While the lowest plasma vitamine C content was determined as 34.54 µg/ml in the control group, it was determined as 53.23, 133.40 and 214.69 µg/ml in the groups receiving rosehip fruit, respectively. When the blood plasma values were examined, the difference among the groups was found to be significant only in terms of triglyceride values in laying hens and uric acid values in roosters ($P < 0.05$). Likewise, the difference among the groups in terms of hatching body weight values of chicks was found to be statistically significant ($P < 0.05$). As a result, it can be said that 5 g/kg rosehip fruit can be added to the diets of breeder hens and roosters exposed to high altitude and cold stress.

Introduction

The significance of rosehip fruit (*Rosa canina*) in terms of health results from its bioflavonoid and carotenoid (lycopene, zeta-carotene, beta-carotene, xanthophyll, neoxanthin and lutein) content in addition to high vitamine and minerals it includes. Beta carotene is the main rosehip carotenoid and it has a significant place in the diet as pro-vitamine A and anti-oxidant (Çınar *et al.*, 2004). In addition to its nutritious value,

rosehip also has protective properties against various ailments and partially treats various diseases (Kadalkal and Nas, 2004). Rosehip fruits are used in the treatment of cold, flu and mild infections (Bown, 1996). It is known that rosehip, which is used as a drug raw material in many European countries, is also used in folk medicine against kidney and bladder stones, diarrhoea, gum bleeding and chest pain (Anonim 2008). Vitamine C, which has a great significance for human health and nutrition, is found in rosehip fruit in highest amount

among fruit types in the world (Ağaoğlu *et al.*, 1987). Rosehip fruits reach the highest vitamin C rate in physiological maturity and this time generally corresponds to September-October (Anonim 2008). Although light coloured and fully ripe rosehip fruits contain more vitamin C, very ripe and dark coloured fruits have less vitamin C (Türkben, 2003). The most important feature of vitamin C is the fact that it destroys the free radicals in our body. Free radicals may cause arteriosclerosis and cancer. Vitamin C is antioxidant due to its property of neutralizing these free radicals (Saraçoğlu, 2006; Untea *et al.*, 2020). In addition to this feature, vitamin C is also used in poultry diets because of its positive effects on the immune system (Sasidhar, 2020) and as an anti-stress factor (Kutlu and Forbes, 1993; Shakeri *et al.*, 2020). Değirmencioğlu and Ak (2003) found that the application of ascorbic acid (0, 50, 100, 150 mg/kg) had no effect on nutrition performance in turkeys fed in the fall period. As a reason, they determined that animals were not exposed to low ambient temperatures. For this reason, it was concluded that new studies conducted in closer regions and at higher doses are needed to show the effects of ascorbic acid more clearly in preventing cold stress in animals.

With 70/524/EEC council directive on feed additives, European parliament and council regulations (EC) 1831/2003 defined CoE 403 numbered *Rosa canina* plant as a natural product and accepted it as a feed additive (Anonymous, 2013). Vitamin C, which is not essential for poultry and which can be synthesized by the animal, is generally not offered in the feed. However, adding vitamin C to the feed is practically applied under stress conditions that reduce the synthesis of vitamin C and increase the need for this vitamin (Kutlu, 2009). Therefore, it is hoped that the oxidative stress that will develop due to cold stress can be reduced with rosehip. The aim of this study was to find out the appropriate usage levels of natural rosehip fruit instead of commercial ascorbic acid in order to meet the vitamin C requirement in breeding hens exposed to cold stress.

Materials and Methods

Animal material of the study consists of 120 Nick Brown hens at the age of 24 weeks and 15 Nick Brown

roosters at the age of 24 weeks which were required for fertilized eggs. After obtaining permission for the study with the 25.10.2018 dated and 2018/10 numbered decision of Van YYU Animal Experiments Local Ethics Committee, the trial phase was carried out in Van YYU Research and Application Farm Directorate coop (at an altitude of 1726 m). 5 groups were formed in the trial. While the group in which no additives were added constituted the control group, the groups in which 100 mg/kg ascorbic acid (vitamin C) and different doses of rosehip fruit (5g/kg, 10g/kg, 15g/kg) were added constituted the treatment groups (Table 1). Each group was divided into three subgroups with 8 hens and 1 rooster with similar average body weights and placed in cages prepared for breeding hens. The rosehip fruit used in the trials was collected in Gevaş and Edremit towns /Van in September and October when the rosehip fruit reached physiological maturity.

Each group was fed with 2850 ME (kcal/kg) basal ration including 16.75% HP prepared for breeding hens (Table 2). The hens and roosters placed randomly in cages were fed with ad-libitum feed and water. During the 12-week trial, 16 hours of light and 8 hours of dark lighting program was applied. In the trial unit, the temperature was set to 24°C for 12 hours and 14°C for 12 hours to create cold stress.

The blood required for checking blood parameters was taken from the under-wing vein. After removing the feathers on the lower surface of the wing, the vein was disinfected; 4-5 ml blood samples were taken into vacuum gel tubes by entering the vein with needle. Plasma was obtained by centrifuging blood samples at 3000 d/min and room temperature for 15 minutes. The plasma was kept at -20°C until analysis. Architect Abbott CI 16200 device and the commercial kits of this device in Van YYU Faculty of Medicine Biochemistry laboratory were used for blood analysis. Vitamin analysis in blood was made in Van YYU Central laboratory.

Nutrient Analysis of Feed

Dry matter, crude ash, crude protein, crude oil, crude fiber nutrient contents of the ration were determined by Wende analysis method (Akyıldız, 1984; AOAC, 1984). Starch, total P, Na, K, Cl and ME values are calculated according to the ration program.

Table 1. Groups in the experiment.

Group 1	Control Group	No Additive
Group 2	Ascorbic Acid (Vitamin C) Group	100 mg/kg Vitamin C
Group 3	Rosehip 1	5 g/kg
Group 4	Rosehip 2	10 g/kg
Group 5	Rosehip 3	15 g/kg

Table 2. Composition and nutrient content of experimental diet.

Raw materials	Rate(g/kg)	Analysed nutrients	(%)
Maize Corn	55.63	Dry Matter	89.62
Feed Flour (46-52)	15.000	Crude Protein	16.75
Soybean Meal (44)	10.967	Crude Cellulose	3.17
Fullfat Soybean	6.346	Crude Oil	3.71
Marble Powder (GRN)	6.326	Crude Ash	10.68
Sunflower Meal (34)	3.032	Starch	44.96
DCP 18	1.636	ME-Pou	11.72
Salt	0.244	ME-Pou	2.80
Sodium Bicarbonate	0.190	Tot-P	0.61
DL-Methionine	0.170	Na	0.16
Vitamine Premix ¹	0.200	K	0.62
Mineral Premix ²	0.100	Cl	0.22
Choline chloride- %60	0.060		
L- Threonine	0.053		
L-Lysine	0.050		

¹: In every 2 kg mixture; 12 500 000 IU Vitamine A, 3 000 000 IU Vitamine D3, 80 000 mg Vitamine E, 5000 mg Vitamine K3, 3000 mg Vitamine B1, 12000 mg Vitamine B2, 55000 mg Niacin, 15000 mg Ca-D-Pantothenate, 4000 mg Vitamine B6, 40 mg Vitamine B12, 2000 mg Folic Acid, 250 mg D-Biotin

²: In every 1 kg mixture; 120000 mg Manganese, 60000 mg iron, 100000 mg zinc, 10000 mg copper, 500 mg Cobalt, 2000 mg iodine, 200 mg Selenium.

Rosehip Vitamine C Analysis

Vitamine C analysis was performed on C₁₈ column (Phenomenex Luna C₁₈, 250 x 4.60 mm, 5 μ) in HPLC (high performance liquid chromatography). Column oven temperature was set to 25°C. Ultra-pure water, pH level adjusted to 2.2 with H₂SO₄, was used as mobile phase in the system at a flow rate of 1 ml/minute. The readings were performed on a DAD detector at 254 nm wave length.

L-ascorbic acid (Sigma A5960) prepared in different concentrations (50, 100, 500, 1000, 2000 ppm) was used to define vitamine C peak and to determine its amount (Demir and Özcan, 2001).

Plasma Vitamine Analyses

Vitamine A, E and C levels in plasma samples were determined with HPLC device. Vitamine A and E analyses were made according to (Zaspel and Csallany, 1983; Miller and Yang, 1985) and vitamine C analyses were made according to (Kartepe, 2004).

Vitamine A and E Plasma Extractions

200 μl plasma was taken into plastic tubes for vitamine A and E analyses. They were added 200 μl ethanol and mixed with vortex for a minute. These were added 800 μl n-hexane and vortexed again for a minute

and centrifuged for 10 minutes at 2000 RPM. 600 μl was taken from the resulting hexane phase and dried under nitrogen gas. The residue was dissolved in 500 μl methanol and injected on the HPLC column (Zaspel and Csallany, 1983; Miller and Yang, 1985).

The setups were made ready for analyses by using vitamine A and E standard. 20 μl was then taken from the prepared extracts and injected into the liquid chromatography column. The diagnoses of vitamine A and E were made using DAD (diode-array detector) detector at 325 and 290 nm wavelengths. Methanol-water (98:2) was used as the mobile phase at a flow rate of 1.5 ml/min. C₁₈ column (4.6 mm x 25 cm) was used to separate the vitamins (Kadikal and Nas, 2004; Donsbough *et al.*, 2010). The calculations were made according to peak area and concentrations of vitamine A and E standards.

Plasma Vitamine C Determination

The levels of vitamine C in plasmas were determined with HPLC-UV method as stated by Kartepe (2004). For plasma analysis, 250 μl of 0.1 M HClO₄ solution was added on 200 μl plasma and vortexed for a few seconds, then 550 μl distilled water was added and after vortexing 10 minutes of centrifugation was made at 4500 RPM. Following these procedures, the supernatant was carefully removed, placed in a vial and vitamine C levels were determined

with HPLC. C18 column (25 cm x 4.6 mm) was used in the HPLC device with a mobile phase of 30 μM KH_2PO_4 methanol (82.5:17.5) at a flow rate of 1.2 ml/min. The readings were made at 250 nm wave length with UV detector. The calculations were made according to peak area and concentrations of vitamin C standards.

Fertility Rate and Chick Quality Values

Fertility rate was found by using the formula below (Türker *et al.*, 2018).

Fertility rate (%) = (The number of fertilized eggs)/(The number of eggs put in machine) X 100

Hatching weight and body length were evaluated as chick quality value. Chick body length was measured with the tip of the chick beak and the tip of the long finger stretched on the ruler (Şeremet, 2012).

Statistical Analysis

In the study conducted according to randomized plot design, SAS (2010) package program was used to analyse the data obtained. DUNCAN multiple comparison test was used to find out the difference between groups (Bek and Efe, 1998).

Results and Discussion

Vitamin C content of the rosehip fruit used in the study was determined as 2862.66 mg in 100 g (Table 3).

Table 3. Vitamin C content of rosehip fruit.

	Vitamin C (mg/100g)
Rosehip fruit	2862.66

This content was found to be higher than the vitamin C content in the rosehip fruit collected in Konya as 2365 mg/100 g and in Kastamonu as 2712 mg/100 g (Demir and Özcan, 2001). This difference can be

attributed to the high altitude of the area and the higher number of sunny days in the area rosehip fruit used in the present study was grown.

When the effect of adding rosehip fruit to the ration on rectal temperature values in roosters was examined (Table 4), the difference between the means of groups was not found to be statistically significant ($P > 0.05$). While the mean rectal temperature of roosters before stress was found as 41.4°C, rectal temperature was found to decrease to 40.9°C with cold stress. It is natural for rectal temperature values of the animals to decrease with cold stress. This difference was not found to be statistically significant in the present study. Similarly, Ahmed *et al.* (2008) reported that the vitamin C supplement added in the drinking water of laying hens raised under sub-tropical conditions did not affect the rectal temperature values significantly. However, Tekeli (2014) added 0 (Control), 10, 20, 30 g/kg rosehip fruit to the ration in a study conducted with broilers and found the pre-stress and post-stress rectal temperature values as 40.8-39.8°C, 40.9-39.7°C, 41.1-39.9°C, 40.8-39.5°C and reported that this difference was statistically significant ($P < 0.05$). This result is not in parallel with the result obtained from the present study. This difference in rectal temperature values may have resulted from the animal's races, ages and differences in the ration they consumed.

In Table 5, when the effect of adding rosehip fruit in the ration on laying hen's fertility rate and the quality values of the chicks was examined, the difference between means of the groups in fertility rate and body length values was not found to be statistically significant ($P > 0.05$). When the egg fertility rates were examined, the highest value was found in the group that received 5g/kg rosehip fruit with 97.53%, while the lowest value was found in the group that received 15g/kg rosehip fruit with 92%. In a study conducted by Shit *et al.* (2012), it was reported that adding vitamin C to the ration of quail exposed to cold stress (10.47°C) had a positive effect on fertility. In the same study, while fertility rate was 72.5% in the control group, it was found as 94.57% in the group which was added 500 ppm L-ascorbic acid.

Table 4. The effect of addition of vitamin C and Rosehip fruit doses to the ration on rectal temperature values in males.

Parameters	Experiment Groups					SEM	P value
	1. Group (Control)	2. Group (Commercial Ascorbic Acid)	3. Group (5g/kg) Rosehip	4. Group (10g/kg) Rosehip	5. Group (15g/kg) Rosehip		
Pre-stress temperature (°C)	41.27	41.50	41.40	41.60	41.36	0.0560	0.2576
Post-stress temperature (°C)	41.00	41.03	40.76	41.00	40.76	0.0617	0.1819

Table 5. The effect of addition of vitamin C and Rosehip fruit doses to the ration on fertility rate and quality values of chicks hatching.

Parameters	Experiment Groups					SEM	P value
	Group 1 (Control)	Group 2 (Ascorbic Acid)	Group 3 (5g/kg) Rosehip	Group 4 (10g/kg) Rosehip	Group 5 (15g/kg) Rosehip		
Fertility rate (%)	94.87	95.12	97.53	95.06	92.00	2.1732	0.8495
Hatching body weight (g)	41.51abc	41.84ab	41.98a	40.23c	40.45bc	0.2086	0.0092
Body length (cm)	17.46	17.20	16.98	16.98	17.11	0.0523	0.0676

SEM: Standard error of difference between means.

*: The difference between the group average shown by different letters on the same line is statistically significant ($P < 0.05$).

In a similar study, it was reported that adding L-ascorbic acid had a positive effect on fertility in poultry under oxidative stress (Ahmadu *et al.*, 2016). The results of these studies are not similar to the results of the present study. This difference can be attributed to the difference in the form and dose of the additives used. The difference between the group means in terms of body weight at hatching was found to be statistically significant ($P < 0.05$).

The highest hatching body weight was found in the group that received 5g/kg rosehip fruit, similar to the fertility rate. In a study on hatching chick weight, it was reported that adding ascorbic acid obtained from 0, 200, 500, 1000 and 1500 mg/kg DM orange peels to the ration did not affect chick hatching weight significantly (Adesola *et al.*, 2013). These results are inconsistent with the results obtained in the present study in terms

of hatching body weight. This difference can be attributed to the environmental conditions of the trial and the differences in forms and doses of the additives used. In addition, factors such as genetic factors, herd age, hatching egg quality, egg collection time, egg storing conditions, incubation temperature and egg weight are also reported to be effective on chick quality features such as chick weight and length (Kamanlı and Durmuş, 2014).

When Table 6 is examined, it was found that the rosehip fruit added in the rations of laying hens which were exposed to cold stress did not affect the amount of retinol and alpha tocopherol in the blood plasma of hens statistically significantly ($P > 0.05$). However, the difference between the groups in terms of the amount of vitamin C in blood plasma was found to be statistically significant ($P < 0.0001$).

Table 6. The effect of addition of vitamin C and Rosehip fruit doses to the ration on laying hens blood plasma vitamin values.

Parameters	Experiment Groups					SEM	P value
	Group 1 (Control)	Group 2 (Commercial ascorbic acid)	Group 3 (5g/kg) Rosehip	Group 4 (10g/kg) Rosehip	Group 5 (15g/kg) Rosehip		
Retinol, ($\mu\text{g/ml}$)	0.28	0.22	0.28	0.26	0.21	0.0112	0.1000
Alpha tocopherol, ($\mu\text{g/ml}$)	4.22	2.93	4.15	3.87	2.84	0.0138	0.138
Vitamin C, ($\mu\text{g/ml}$)	34.54c	47.06c	53.23c	133.40b	214.69a	6.4326	<0.0001

SEM: Standard error of difference between means.

*: The difference between the group average shown by different letters on the same line is statistically significant ($P < 0.05$).

While the plasma vitamin C value was 34.54 µg/ml in the control group, the highest vitamin C value was found in the group that received 15 g/kg rosehip fruit with 214.69 µg/ml. Similarly, Ahmed et al. (2008) reported that levels of vitamin C added in increasing levels in the drinking water of laying hens placed under sub-tropical conditions increased plasma vitamin C levels significantly. Exogenous vitamins such as alpha tocopherol and vitamin C protect the cells against lipid peroxidation.

Lipid peroxidation leads to the deterioration of physiological functions including immunity, growth and reproduction (Altiner *et al.*, 2017). The linear increase in the vitamin C level in blood with increasing doses of rosehip fruit can be explained with adding the rosehip fruit in the ration in increasing doses.

As can be seen in Table 7, the rosehip fruit added in the rations of laying hens exposed to cold stress significantly affected the level of triglyceride in hens' blood plasma ($P < 0.05$). In the present study, while the level of triglyceride in the control group was 1214.00 mg/dL, it was found to decrease significantly in the group that received 5 g/kg rosehip fruit and it was found as 409.60 mg/dL. High level of triglyceride in the blood has been associated with the emergence of a large number of important diseases, mainly cardiovascular diseases (Tada *et al.*, 2018). Therefore, 5g/kg rosehip supplement is evaluated as important in terms of health. Similarly, Mutlu et al. (2015) reported that gypsum extract decreased blood triglyceride level significantly in quails exposed to cold stress. Behboudi et al. (2016) reported that the use of lemon juice as a source of vitamin C significantly lowered the level of triglyceride in broilers bred under heat stress. Unlike the present study, in their study they added *Berberis vulgaris* fruit used as a source of vitamin C in laying hens' rations, Kermanshahi and Riasi (2006) reported that the value of

blood triglyceride was not significantly affected. Tekeli (2014) reported that using different doses of rosehip fruit in broilers exposed to cold stress did not affect blood plasma triglyceride levels significantly. Arpat (2016) reported that rosehip fruit used in laying hens did not have a significant effect on blood triglyceride value. The inconsistency between these studies may have resulted from the differences in the source of vitamin C, type of animal, sex of the animal, purpose of breeding, conditions of breeding and the content of the basic ration used. As can be seen in Table 7, the rosehip fruit added in the rations of laying hens exposed to cold stress caused numerical differences in the amount of cholesterol, glucose, uric acid, sodium, ALT, AST and GGT in hens' blood plasma, while this difference was not found to be statistically significant ($P > 0.05$). Similarly, Tekeli (2014) reported that rosehip fruit added in the ration of broilers did not affect the glucose and uric acid levels. Arpat (2016) reported that adding different rates of rosehip (0, 0.5, 1, 2, 4 and 8%) in the rations of laying hens did not affect blood plasma, cholesterol, AST and ALT values.

Unlike the present study, in a study conducted on broilers, it was reported that adding rosehip fruit to different doses of ration (0, 10, 20, 30 g/kg) under cold stress affected the level of blood plasma cholesterol level significantly (Tekeli, 2014). In their study they added *Berberis vulgaris* fruit to the rations of laying hens as 0, 0.5, 1, 1.5 and 2% vitamin C Kermanshahi and Riasi (2006) found that the total cholesterol amount decreased significantly ($P < 0.05$). In the present study (Table 7), it was found that blood plasma GGT values were numerically lower in all treatment groups. Low GGT value is evaluated as positive in terms of liver and animal health (Kale, 2019).

Table 8 shows the effects of vitamin C and rosehip fruit added in roosters' ration on blood plasma values.

Table 7. The effect of addition of vitamin C and Rosehip fruit doses to the ration on blood plasma values in laying hens.

Parameters	Experiment Groups					SEM	P value
	Group 1 (Control)	Group 2 (Commercial ascorbic acid)	Group 3 (5g/kg) Rosehip	Group 4 (10g/kg) Rosehip	Group 5 (15g/kg) Rosehip		
Cholesterol (mg/dL)	121.00	106.40	145.20	110.33	81.25	10.6112	0.3695
Glucose (mg/dL)	223.00	232.00	212.20	235.50	239.75	3.6209	0.0942
Triglyceride (mg/dL)	1214.00a	980.33ab	409.60c	1163.83a	688.25bc	69.3159	0.0019
Uric Acid (mg/dL)	4.08	3.78	4.80	4.82	5.55	0.3526	0.4711
Sodium (mmol/L)	157.80	149.00	148.20	147.67	151.00	4.5826	0.4434

It was found that the vitamin C and rosehip fruit added in roosters' ration caused numerical differences in the amount of cholesterol, glucose, triglyceride, sodium, ALT, AST and GGT in roosters' blood plasma; however, this difference was not found to be statistically significant ($P > 0.05$). In terms of the amount of uric acid in blood plasma, the difference between the control group and the other groups was found to be statistically significant ($P < 0.05$). As can be seen in Table 7 and Table 8, while the rosehip supplement caused a difference only in triglyceride levels of laying hens, it caused statistical difference only in uric acid in roosters ($P < 0.05$).

In roosters, uric acid level which was 11.03 mg/dL in the control group decreased in groups in which rosehip was added and it was found as 7.66, 8.30, 8.50 and 8.16 mg/dL, respectively. Donsbough *et al.* (2010) reported that serum uric acid level could be used as an indicator of amino acid availability in broilers fed with sufficient and insufficient rations in terms of amino acid level. High uric acid level in plasma is considered as a risk factor for gout, renal diseases, metabolic syndrome and cardiovascular diseases (Oliveira and Burini, 2012).

Conclusion

When compared with high doses, low doses of rosehip fruit had a positive effect on hatching live weight; increased plasma vitamin C content when compared with the control group and the group which was given commercial ascorbic acid supplement; reduced the level of triglyceride and reduced the level of uric acid in roosters. Due to these results, it can be recommended to add 5 g/kg rosehip fruit instead of commercial ascorbic acid in the rations of laying hens and roosters exposed to high altitude and cold stress in order to meet their vitamin C need. Using rosehip fruit as a feed additive in livestock will be a great benefit to our country's economy in terms of utilizing our natural resources. New comprehensive studies including sperm and all incubation parameters are needed to fully reveal the effect of rosehip fruit on breeding animals exposed to cold stress.

Conflict of Interest

The authors declare no conflict of interest.

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